



# The effectiveness of HEPA filters to capture DNA

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# Presentation agenda



Types of BSCs and  
HEPA filters



Why do we wonder  
about DNA?



Experimental Design



Results



Conclusions



Recommendations

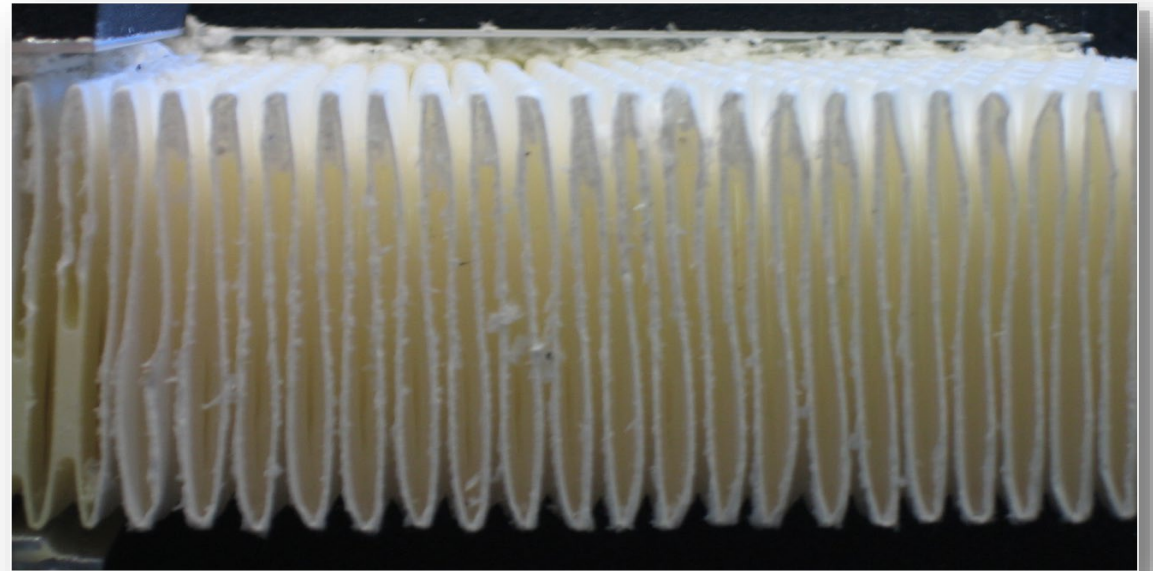
# Biosafety Cabinets (BSCs)

- A ventilated enclosure for work with biohazard agents assigned to biosafety levels 1 through 4, as per the BMBL.
- Provides 3 types of **CONTAINMENT**:
  - Personnel protection
  - Product protection
  - Environmental protection



# HEPA filtration

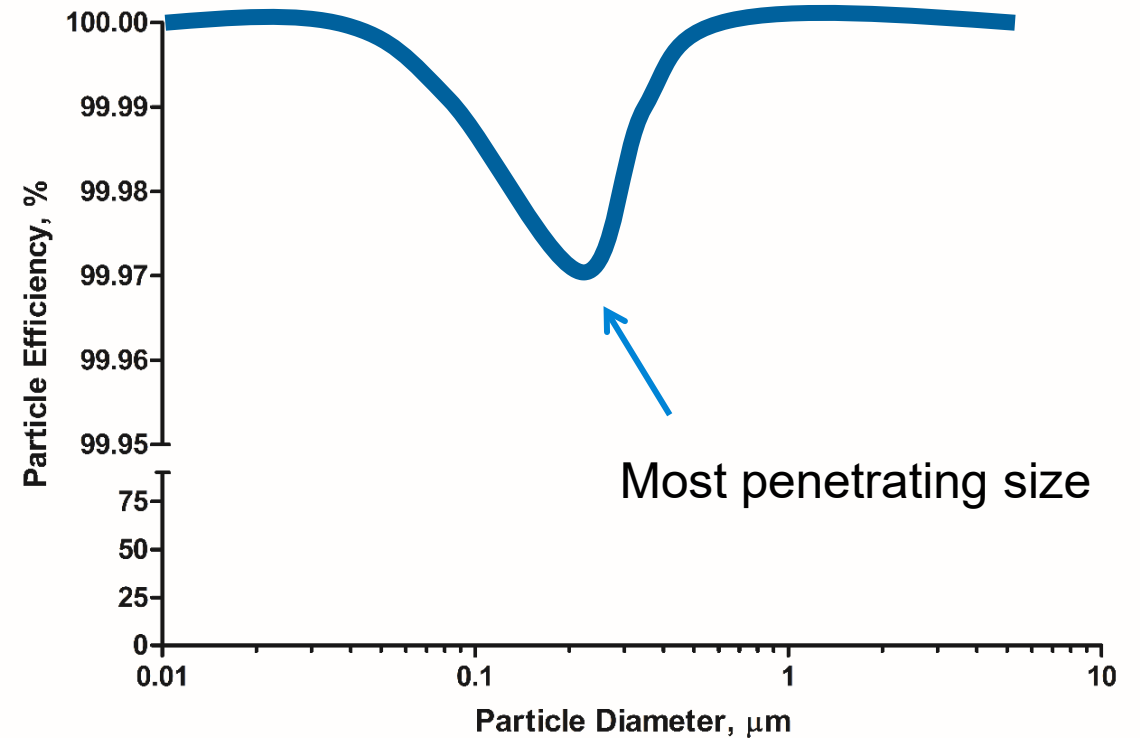
- **H**igh **E**fficiency **P**articulate **A**ir
  - Filter out 99.97% (or better) of 0.3  $\mu\text{m}$  mass median particles of DOP (Dispersed Oil Particulate)
  - **CANNOT filter out gasses and vapors!**



TEST-RP-CC-001

# HEPA filtration

- The **worst** filtration occurs at the “most penetrating size” which is  $0.22\mu\text{m}$ . Smaller and larger particles are captured **better**.



# HEPA filtration in a BSC

NSF International Standard 49 requires HEPA filters used in BSCs to be a minimum of **99.99% effective** when tested with 0.3µm particulates.

# Biosafety Cabinet Classification

## Class I

Personnel and Environmental Protection

## Class II

Personnel, Product, and Environmental Protection

## Class III

For highly infectious agents  
Gas-tight, physical barrier

### Type A1

Recirculated  
Minimum 75 fpm intake velocity

### Type A2

Recirculated  
Minimum 100 fpm intake

### Type B1

Partial recirculation  
Minimum 100 fpm intake

### Type B2

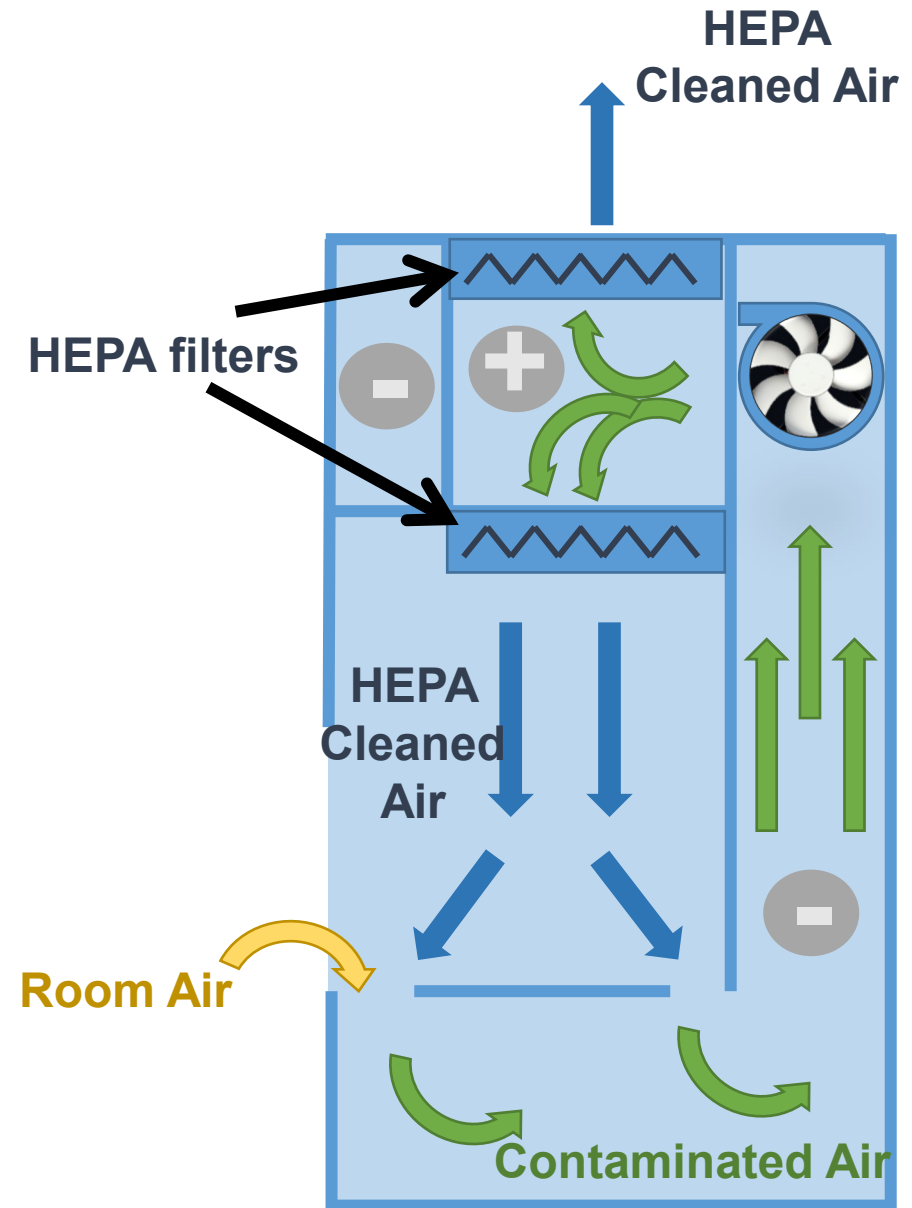
100% Exhaust  
Minimum 100 fpm intake

### Type C1

Partial recirculation  
Minimum 100 fpm intake

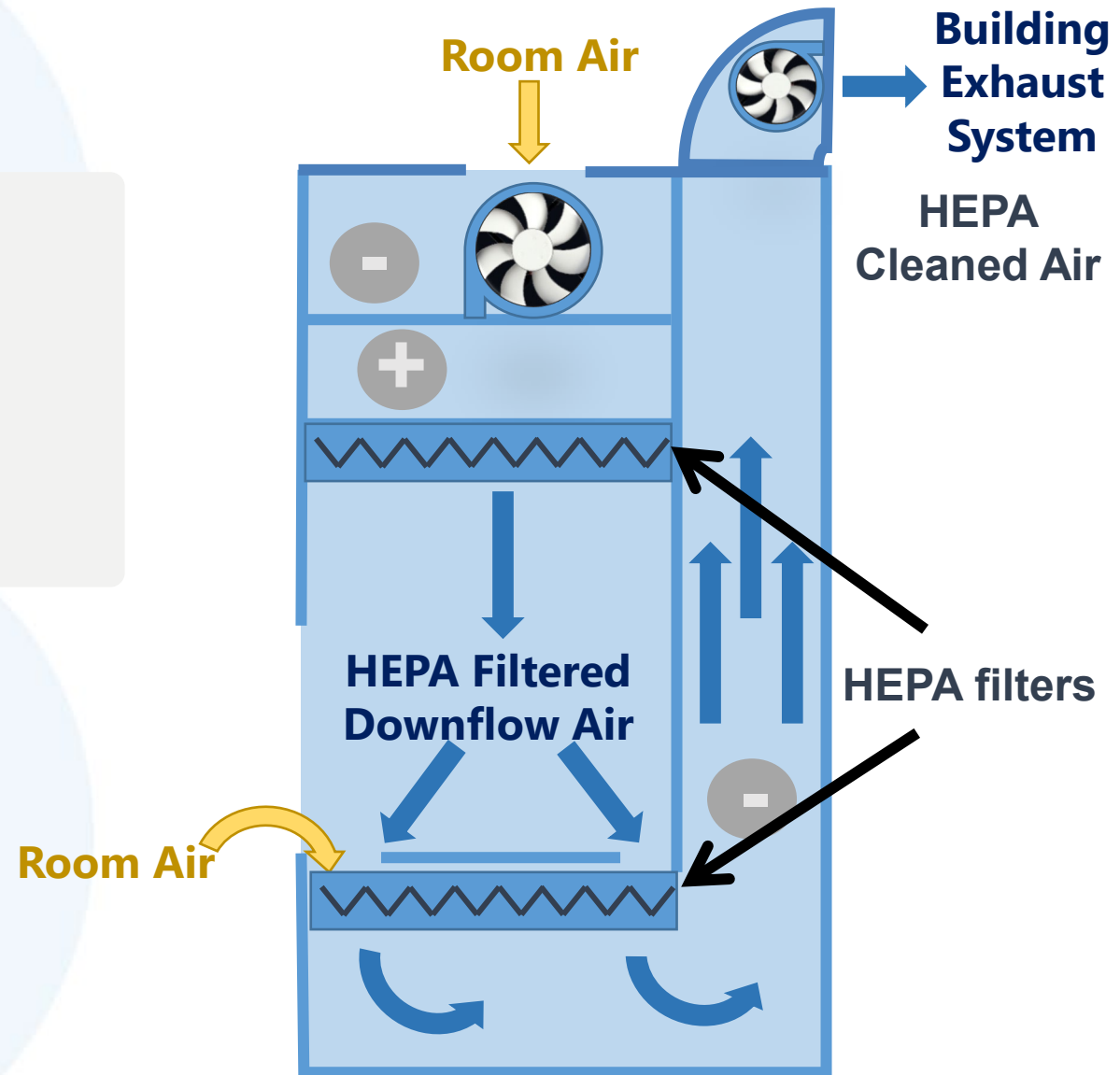
# Class II Type A2 BSCs

- Personnel, Product and, Environmental Protection
- Minimum 100 fpm intake air
- Partial recirculation



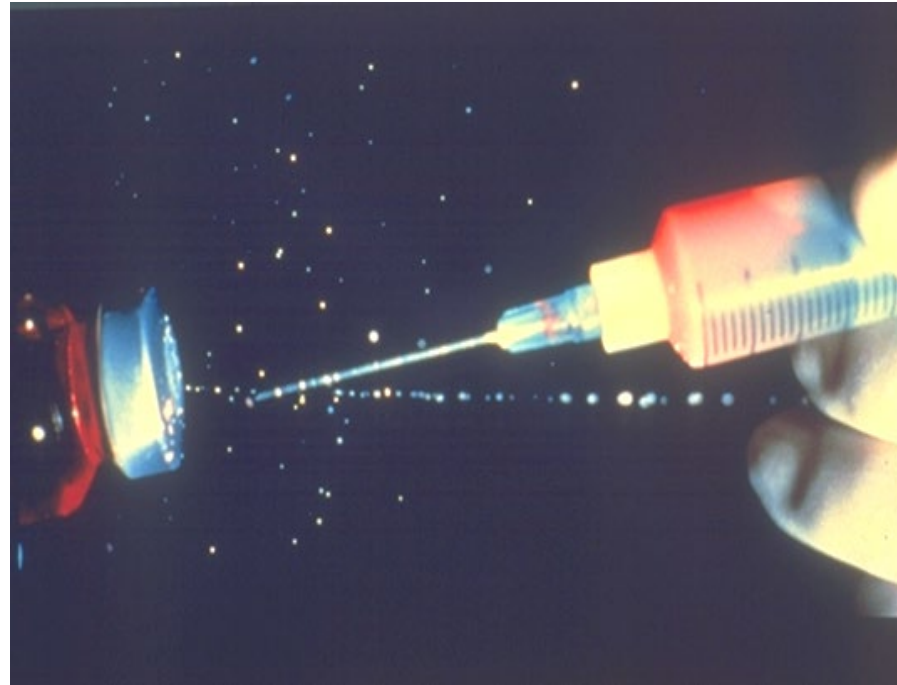
# Class II Type B2 BSCs

- Personnel, Product and, Environmental Protection
- Minimum 100 fpm intake air
- 100% Exhausted



# Aerosols

- Many laboratory procedures create aerosols during routine use.
  - Syringes
  - Centrifuges
  - Pop cap tubes
  - Blowing out pipettes
  - Pouring liquids
  - Shaking or vortexing tubes



# Is DNA captured in a BSC?

- DNA is small
- “Sticky” due to negatively charged backbone
- “Special” molecule
- Does it become dislodged from HEPA?
- Currently UNKNOWN



# Experimental Design

- Test #1: Direct application of DNA aerosol to HEPA filter
  - Can DNA be detected downstream of the HEPA filter? If so, how much?

- Test #2: DNA aerosol within a Class II Type A2 BSC (Internally recirculating)
  - Is DNA detected on the internal walls of the BSC?
  - Does it pass through the HEPA filters? If so, how much?
  - Can it be removed with a standard fumigation or bleach and ethanol?

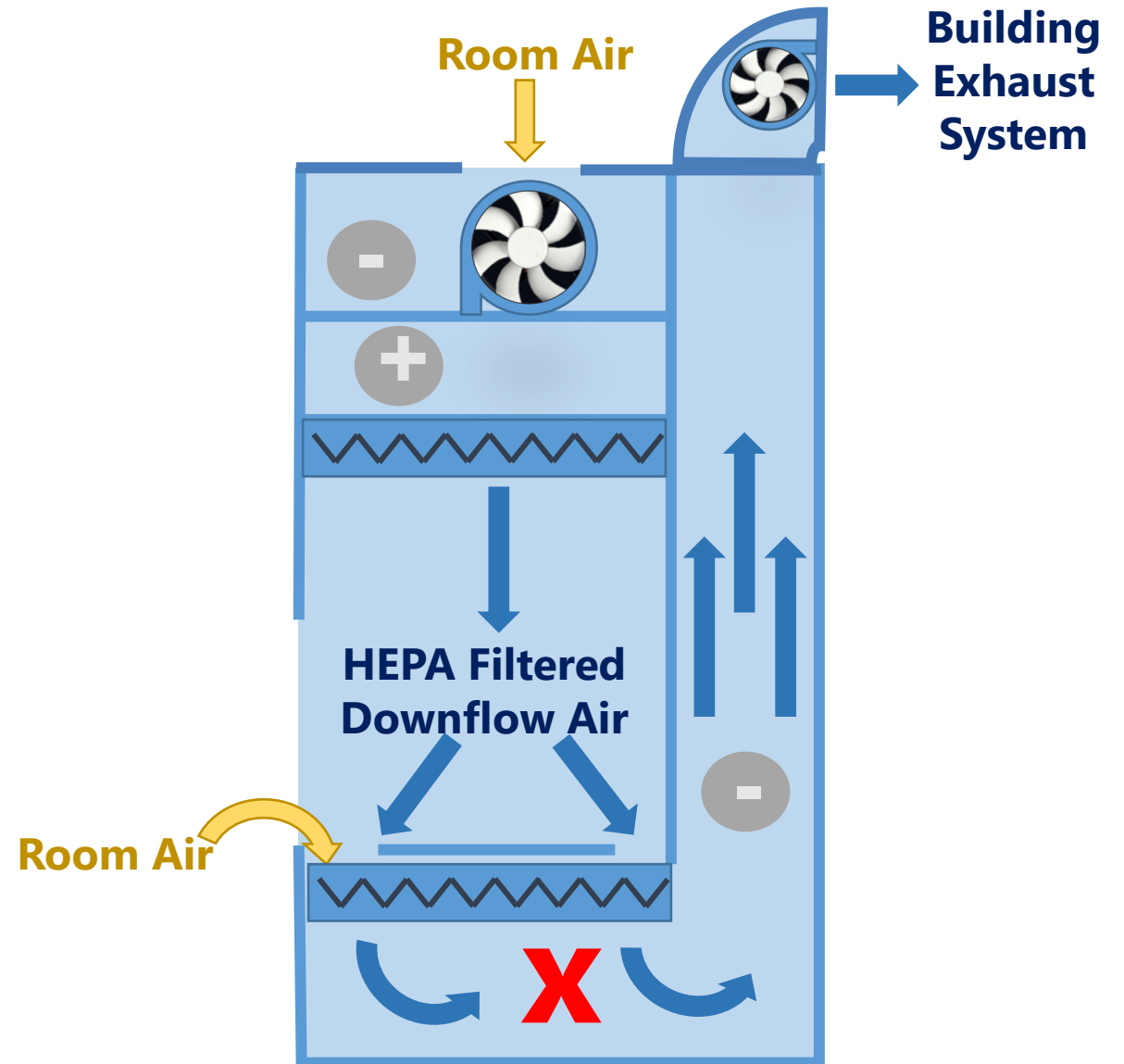
# Creating DNA aerosol

- Collision Nebulizer
  - Creates  $\sim 1\mu\text{m}$  droplets
- $10^9$  copies DNA/mL TE buffer
  - Known amount of DNA applied to BSC



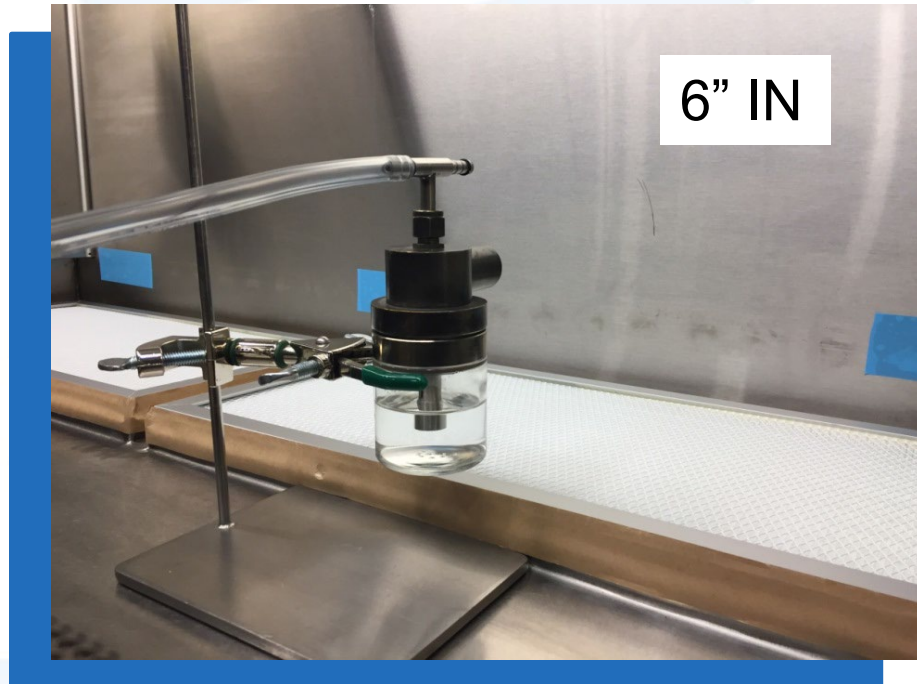
# Baker's Class II Type B2 BSC – BioChemGARD

- Remove the worksurface to use the exhaust HEPA filter
- Power Failures to the Building Exhaust system can be simulated
- Filters are easy to remove
- X = sample location



# Experimental Setup

- Nebulizers were placed 6, 9, and 12 inches above the HEPA filters
- Experiments tested with filters in place (IN) and removed (OUT)



# Test #1: Direct Application Results

| Filter IN/OUT | Location | Avg Copies of DNA detected |
|---------------|----------|----------------------------|
| IN            | Left     | 0                          |
| IN            | Right    | 0                          |
| OUT           | Left     | $2.5 \times 10^7$          |
| OUT           | Right    | $3.3 \times 10^7$          |
| Power Failure | Left     | 0                          |
| Power Failure | Right    | 0                          |

# Summary, Part 1

DNA aerosols applied directly to a HEPA filter are captured with 100% efficiency

Power Failures and Mechanical forces will not dislodge the DNA from the HEPA filter.

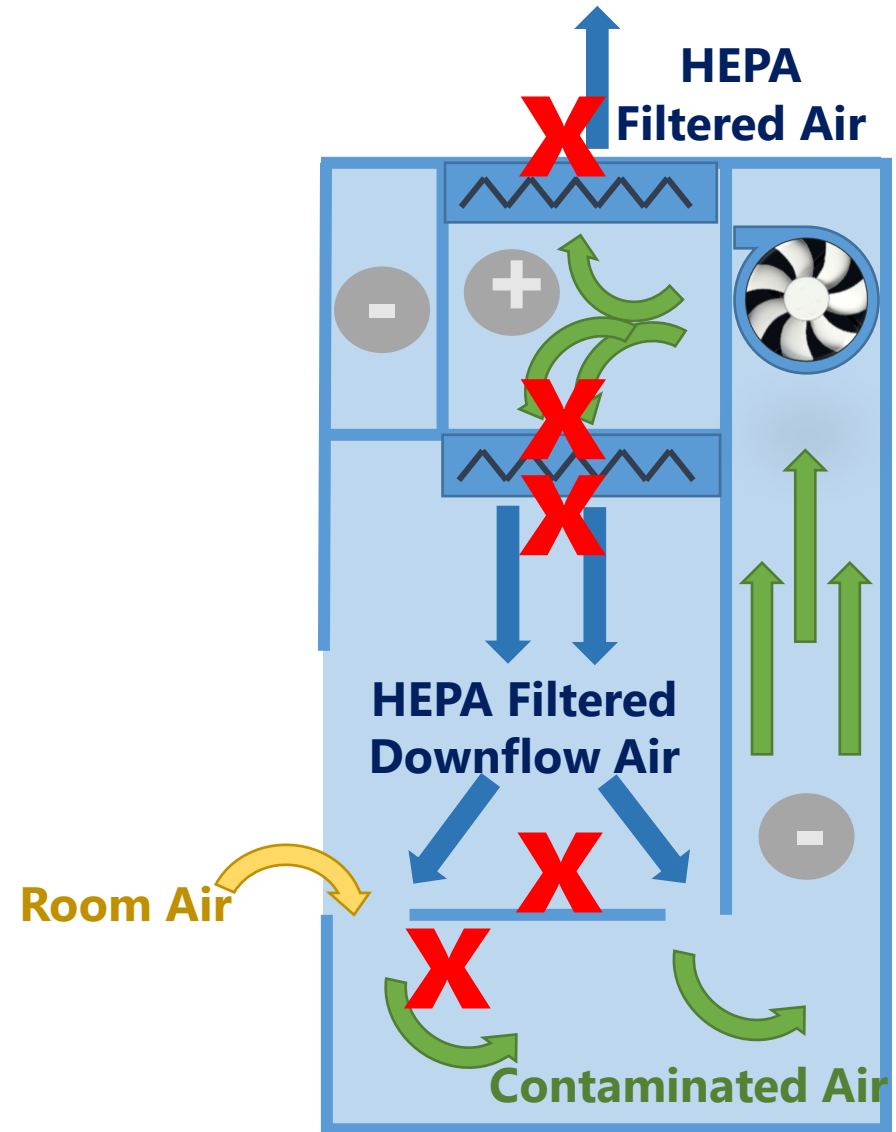
# Experimental Design

- Test #1: Direct application of DNA aerosol to HEPA filter
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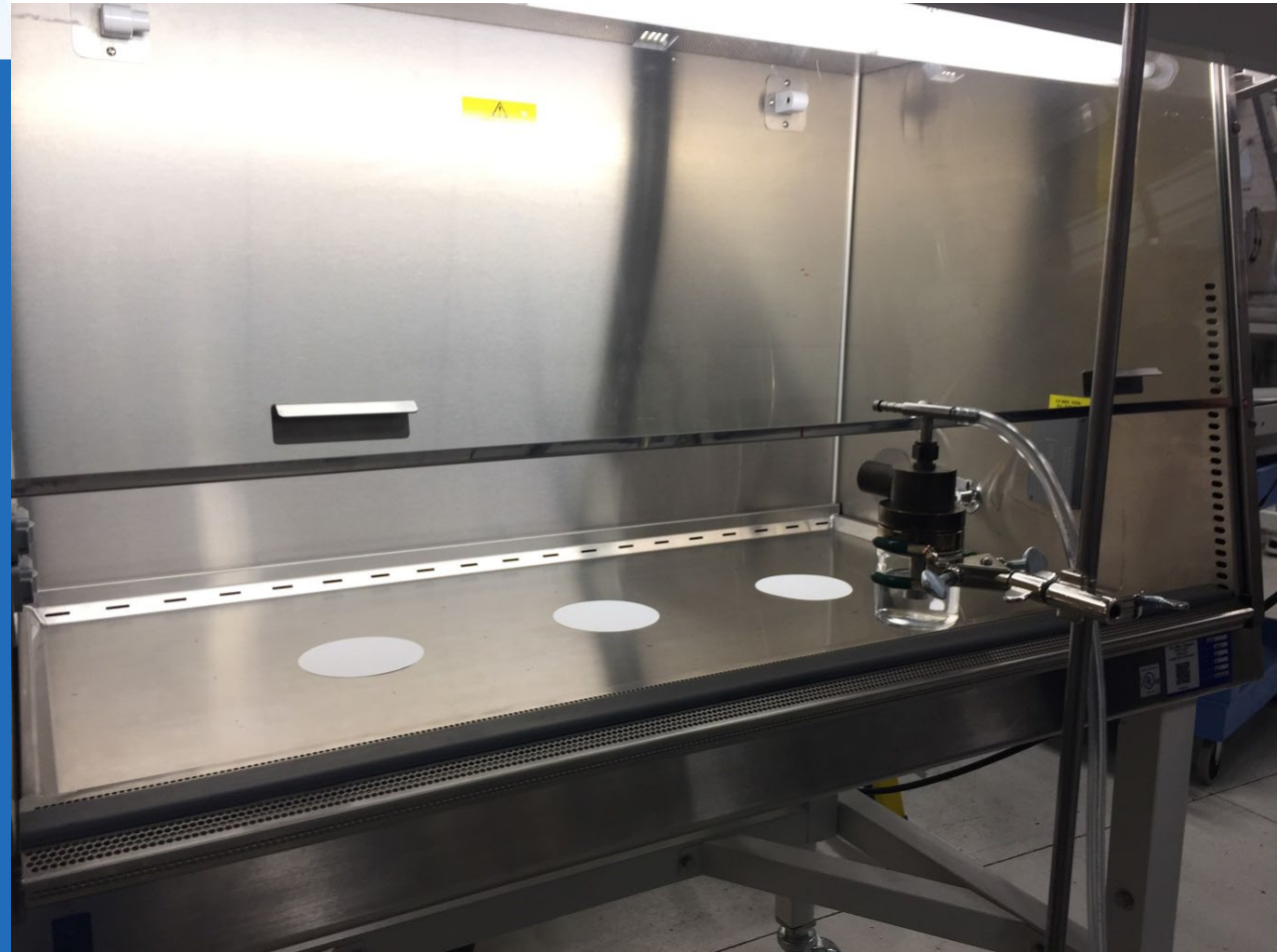
# Baker's Class II Type A2 BSC – SterilGARD

- Partially internally recirculating
- May be vented back to the room or outdoors with a canopy connection
- X = sample location



# Nebulizer placement

- Samples taken by Whatman filter paper and cotton swabs on the stainless steel surface or glass microscopy slides



## Test #2: DNA in a BSC Results

| Location                | Avg Copies of DNA detected | HEPA Capture efficiency (%) |
|-------------------------|----------------------------|-----------------------------|
| Pre-cleaning            | 0                          |                             |
| Under worksurface       | $2.17 \times 10^7$         |                             |
| Worksurface             | 65                         | 99.9997%                    |
| Upstream Supply HEPA    | $6.25 \times 10^4$         |                             |
| Downstream Supply HEPA  | 159                        | 99.9991%                    |
| Downstream Exhaust HEPA | $2.37 \times 10^3$         | 99.9945%                    |

# DNA removal

The DNA contaminated BSC was tented and subjected to chlorine dioxide fumigation overnight

Bleach (10%) and ethanol (70%) were wiped on the contaminated surface for DNA removal as well

## Test #2: DNA removal efficiency

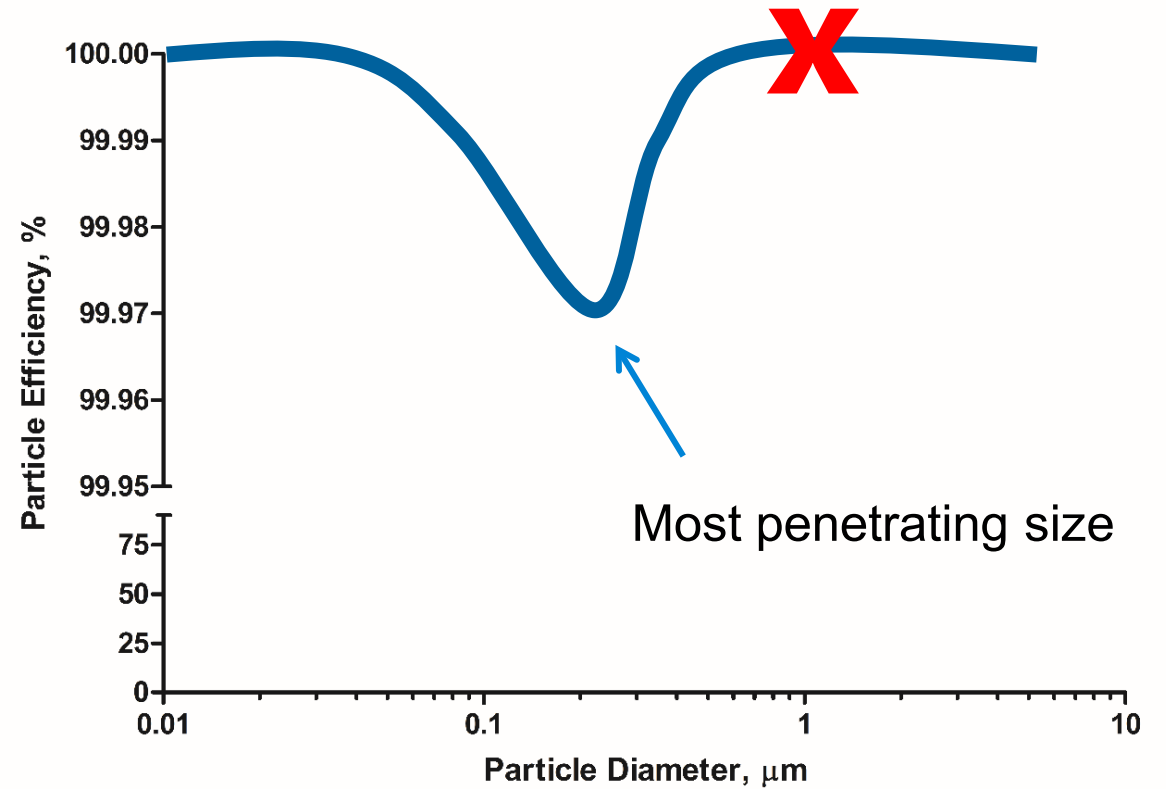
| Location             | Avg Copies of DNA detected | Removal efficiency (%) |
|----------------------|----------------------------|------------------------|
| Pre-decontamination  | $3.31 \times 10^9$         |                        |
| Post-decontamination | $1.07 \times 10^7$         | 99.7%                  |
| Post-cleaning        | 0                          | 100%                   |

# Summary

- DNA aerosols applied directly to a HEPA filter are captured with 100% efficiency
- Power Failures and Mechanical forces will not dislodge the DNA from the HEPA filter.
- DNA within the airstream of a BSC is removed with 99.99+% efficiency. Some cross-contamination is possible.
- Chlorine Dioxide decontamination only removes 99.7% DNA. Bleach and ethanol remove 100%.

# So, what's going on?

- 1 $\mu\text{m}$  droplets are captured 100%
- As the droplets pass through the air, they dry becoming smaller (0.2 $\mu\text{m}$ )
- This reduces capture efficiency.



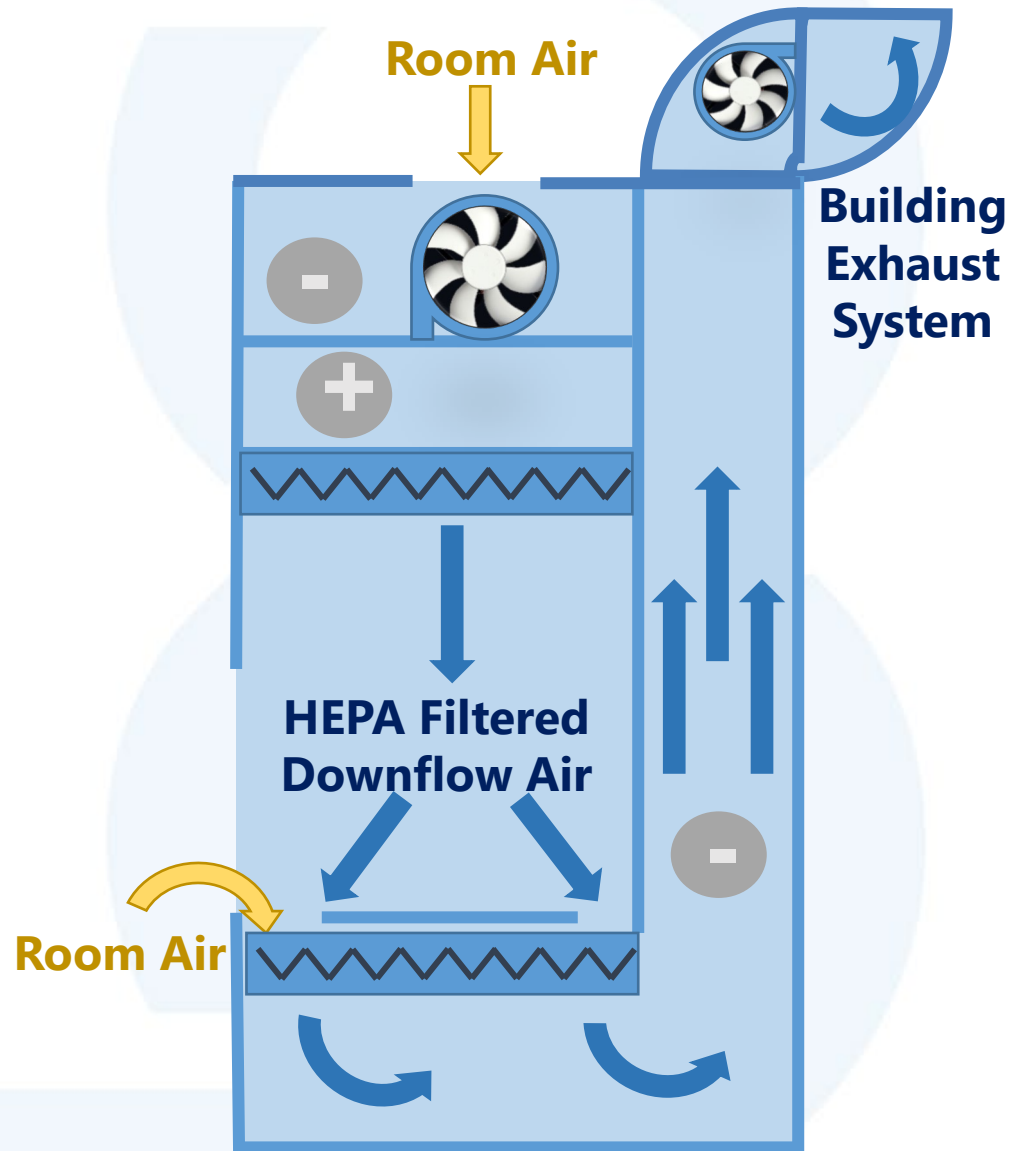
# Conclusions

- DNA aerosols may be captured by HEPA filters
  - Size dependent
- DNA dried particles will be removed according to the HEPA's efficiency rating.
- Fumigation decontamination is not enough to remove all DNA. Bleach, ethanol, and physical wiping are required.

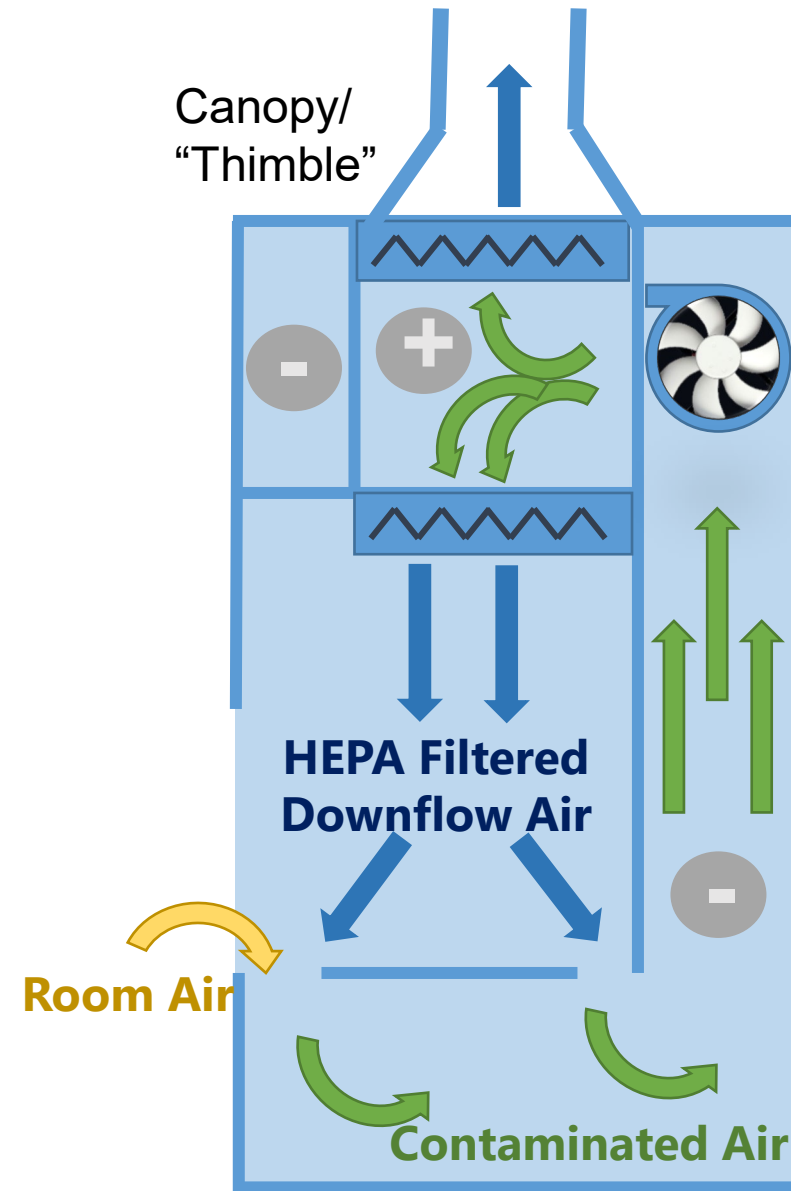
# Recommendations

- Use a higher efficiency rated HEPA filter
  - ULPA = 99.999%
  - SULPA= 99.9999%
- BSC selection is critical, dependent on where your contamination concern is

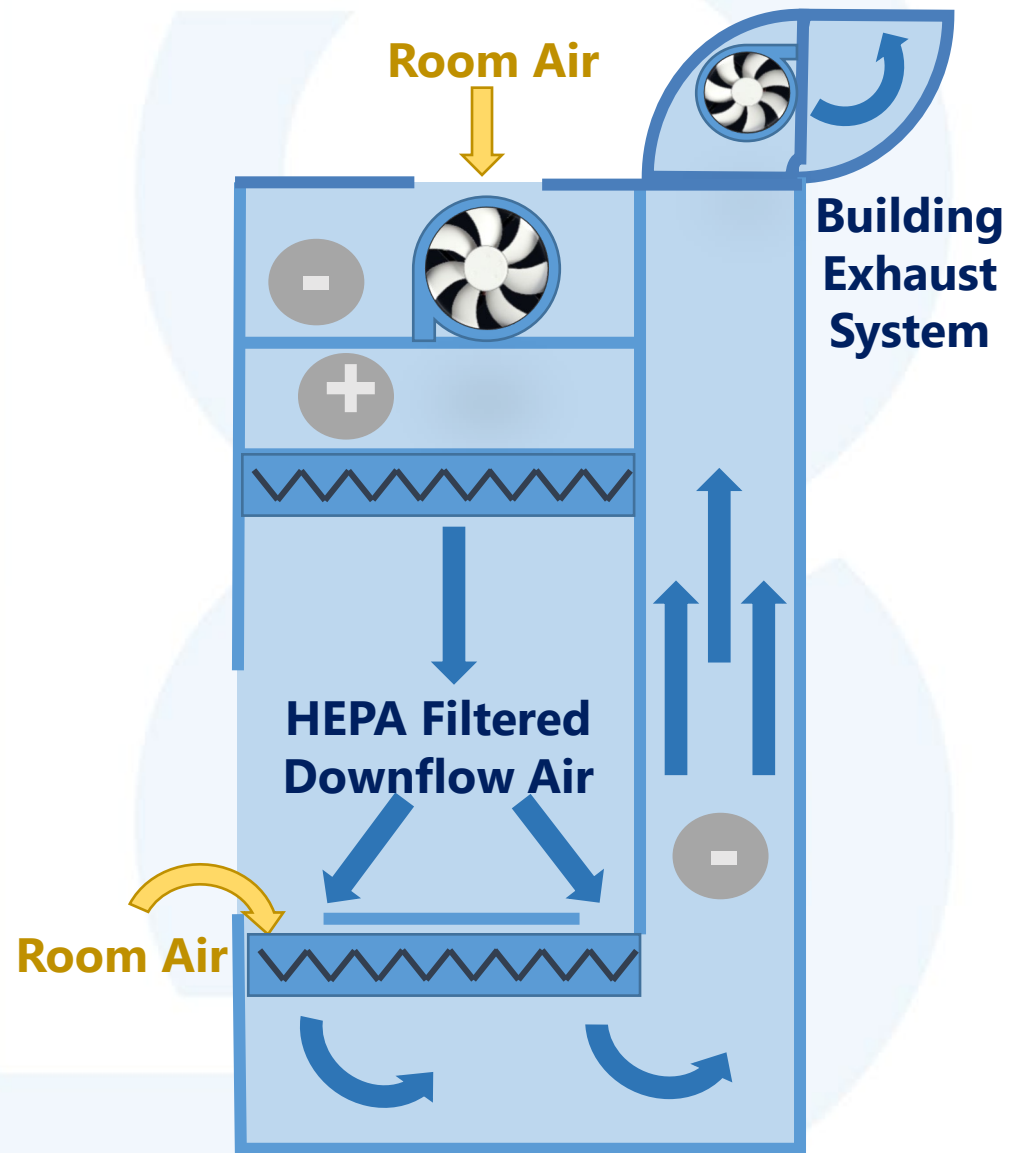
## B2 – BioChemGARD



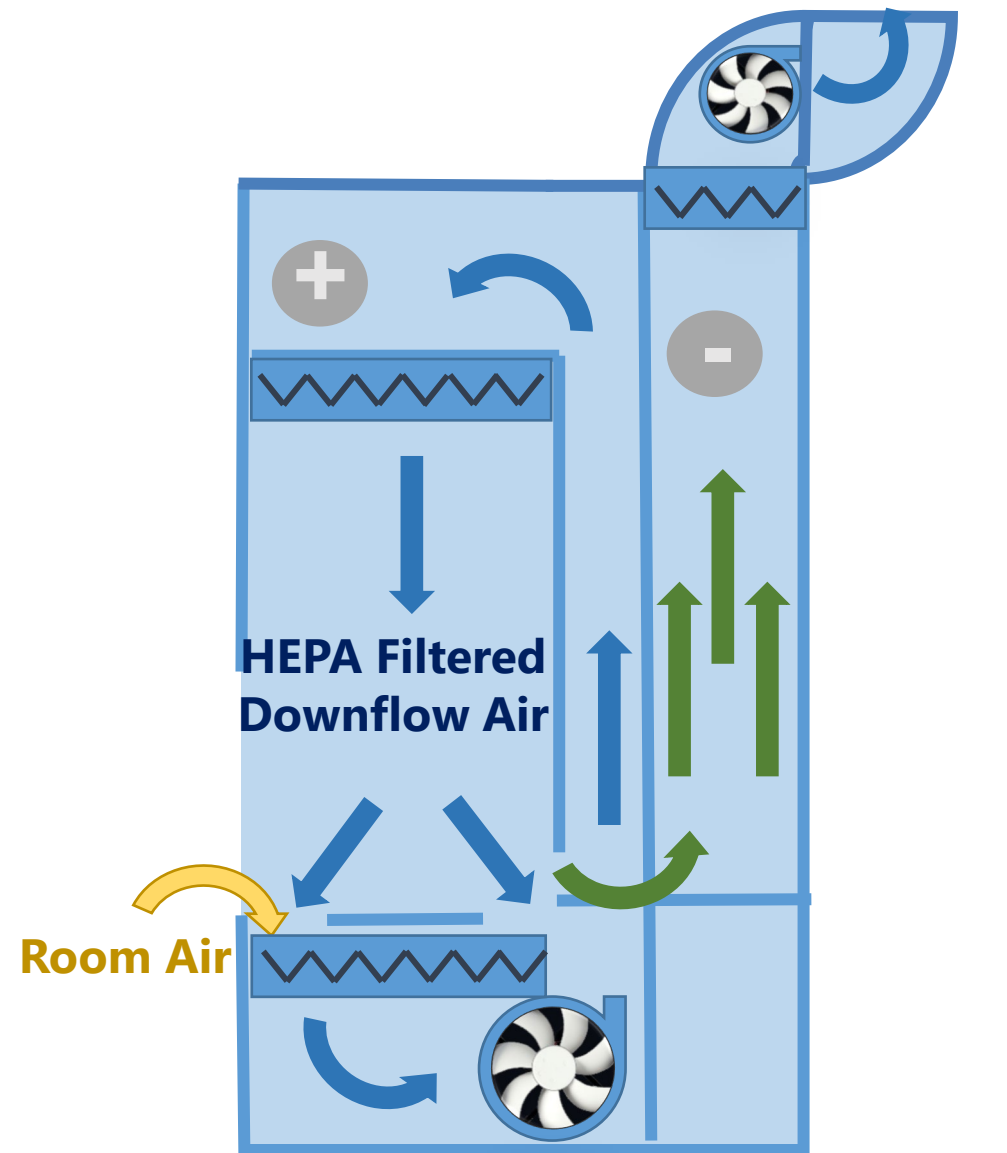
## A2– SterilGARD



**B2 – BioChemGARD**



**B1– NCB**



# Acknowledgments

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# References

- Held, K.F., Rundell, C., Thibeault, R., Ghidoni, D., Magoon, D., Nguyen, T., Fisher, A., Parker, B., Spenlinhauer, T., Gordon, J., and Nesbitt, S. (2018) The Effectiveness of HEPA Filters on DNA. *Applied Biosafety*. Vol. 23(2) 1-5.
- NSF International. (2016) NSF/ANSI 49 – 2016 Biosafety Cabinetry: Design, Construction, Performance, and Field Certification. Ann Arbor, MI.
- Tsoi, M., Do, T.T., Tang, V., Aguilera, J.A., Perry, C. C., and Milligan, J.R. (2010) Characterization of condensed plasmid DNA models for the studying the direct effect of ionizing radiation. *Biophys Chem*. 147(3): 104-110.
- May, K. R. (1973) The Collison Nebulizer: Description, Performance and Application. *Aerosol Science*. 4: 235-243.
- Stuart, D.G., First, M.W., Jones, Jr., R.L., and Eagleson, J.M. (1983) Comparison of Chemical Vapor Handling by Three Types of Class II Biological Safety Cabinets. *Particulate & Microbial Control*. 2:18-24.
- Held, K.F., Ghidoni, D., Hazard, G., and Eagleson, D. (2016) Volatile Chemicals and Class II Type A2 “Recirculated” BSCs: How Much is Safe? The Baker Company.



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